

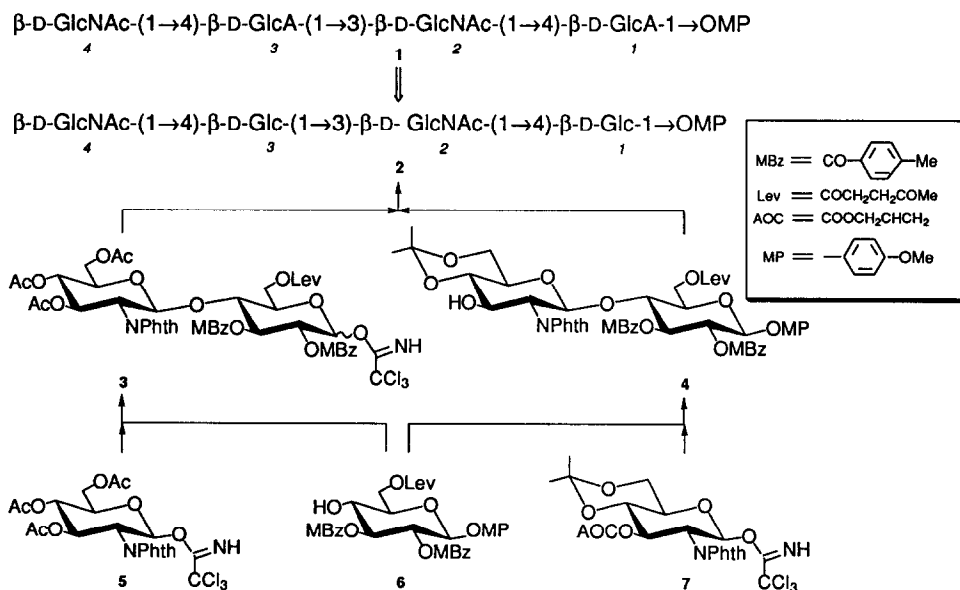
SYNTHESIS OF A TETRASACCHARIDE FRAGMENT OF HYALURONIC ACID HAVING A GLUCURONIC ACID AT THE REDUCING END¹

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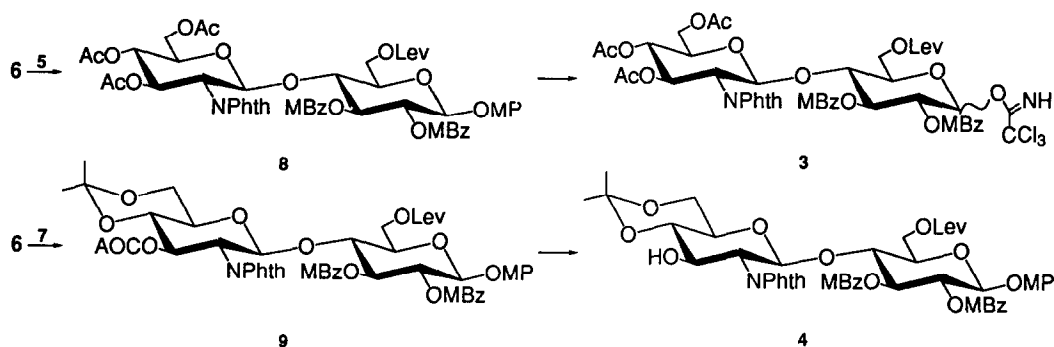
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Abstract: A stereocontrolled synthesis of a tetrasaccharide fragment of hyaluronic acid, β -*p*-methoxyphenyl glycoside of β -D-GlcNAc-(1 \rightarrow 4)- β -D-GlcA-(1 \rightarrow 3)- β -D-GlcNAc-(1 \rightarrow 4)-D-GlcA, is presented.

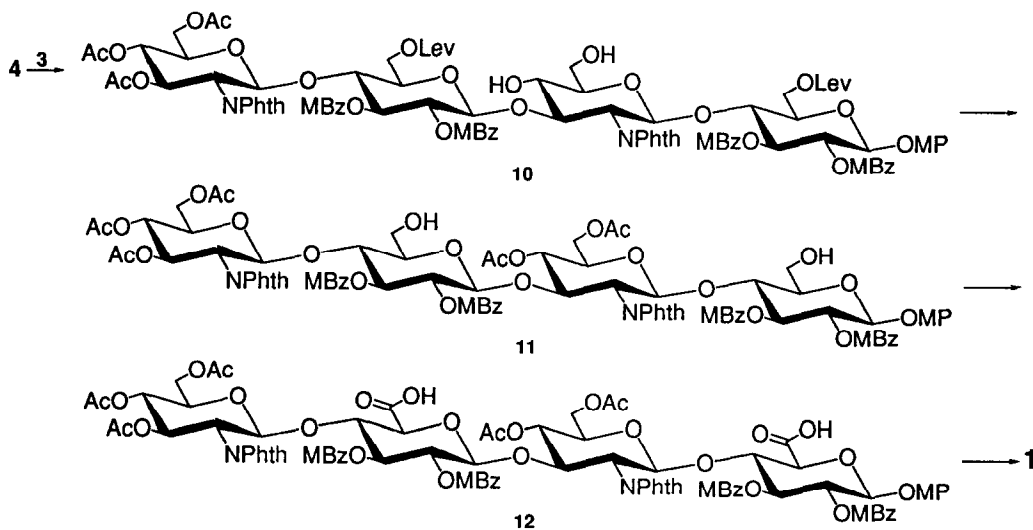
Hyaluronic acid (HA) is a linear, extracellular carbohydrate polymer consisting of disaccharide repeating units², namely $[\rightarrow 4)\text{-}\beta\text{-D-GlcpA-(1}\rightarrow 3)\text{-}\beta\text{-D-GlcpNAc-(1}\rightarrow n]$. It is a major component in extracellular matrices³ and regulates various biological processes such as cellular proliferation^{4,5}, cell-cell recognition³, cell migration⁶, and cell adhesion. Interestingly, high concentrations of HA suppress vascularisation⁷, while enzymically (hyaluronidase) generated medium-sized fragments of HA oligosaccharides at low concentrations stimulate the formation of new capillary bloodvessels^{8,9}. Recently, it was suggested that higher concentrations of oligosaccharides of HA suppress the initial growth, and therefore it was postulated that they might act as a potential antitumor drug¹⁰.



Isolation of the various oligosaccharides is laborious and so far mainly mixtures of even numbered oligosaccharides were used in biological testing^{8,9}. The finding that these oligosaccharides can modulate angiogenesis initiated a synthetic program for the preparation of a wider range of medium-sized oligosaccharide fragments built up of even or odd number of monosaccharides with GlcA or GlcNAc at the reducing end. This report describes the stereoselective synthesis of HA fragment 1. Oxidation of the primary hydroxyl groups (2) was carried out in the final stage of the synthesis since the C-4 hydroxyl group of D-glucuronic acid has a low reactivity towards glycosidation¹¹. Therefore, key tetrasaccharide 2 was designed, and synthesised from glycosyl donor 3 and glycosyl acceptor 4. These synthons can be prepared by coupling of glycosyl donor 5 with glycosyl acceptor 6 and glycosyl donor 7 with 6, respectively.



The necessary monosaccharide building units 5, 6, and 7 were synthesised as described earlier^{1,12}. Stereocontrolled glycosylation of 6¹³ with 1.5 equivalents of 5 in CH₂Cl₂ in the presence of BF₃·OEt₂ and molecular sieves 4A at 25°C afforded in 81% 8¹³, which was converted into glycosyl donor 3¹³ in 2 steps (*I* CAN¹⁴ in 1:1:1.4 toluene-H₂O-MeCN, 2 CCl₃CN¹⁵, DBU in CH₂Cl₂, 88% overall).



Also stereocontrolled condensation of **6** with 1.5 equivalents of **7** in CH₂Cl₂ in the presence of BF₃·OEt₂ and molecular sieves 4A gave **9**¹³ in 81% yield, which was subsequently treated with (Ph₃P)₄Pd^{16,17} and morpholine in THF to afford in 88% glycosyl acceptor **4**¹³. TMSOTf-MS4A promoted glycosylation of **4** with disaccharide donor **3** (2.5 equivalents) in CH₂Cl₂ at 0°C gave after acid hydrolysis with TFA in CH₂Cl₂ and water tetrasaccharide **10**¹³ in 77% yield. Compound **10** was further converted into **11** in 2 steps (1 Ac₂O-DMAP in py, 2 NH₂NH₂·HOAc in 2:1 EtOH-toluene^{18,19}, 85% overall). The oxidation of the two primary hydroxyl groups to obtain **12**¹³ was successfully achieved in 2 steps (1 DMSO, (COCl)₂, iPr₂NEt²⁰, -78°C, 2 NaClO₂, NaH₂PO₄ in 3:2:1 *t*BuOH-H₂O-2-methylbutene^{21,22}, 76% overall). The final deprotection of **12** into **1** was carried out in 2 steps (1 MeNH₂²³ in MeOH, 2 Ac₂O in MeOH, 59% overall).

In summary, a stereocontrolled synthesis of HA tetrasaccharide **1** was carried out in a highly efficient manner by using one monosaccharide acceptor **6** and two monosaccharide donors **5** and **7**.

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References and Notes

- Part 3. For part 2, see T.M. Slaghek, Y. Nakahara, T. Ogawa, J.P. Kamerling, and J.F.G. Vliegthart, *Carbohydr. Res.*, submitted. For part 1, see: T.M. Slaghek, Y. Nakahara, and T. Ogawa, *Tetrahedron Lett.*, **33**, 4971 (1992).
- K. Meyer, *Fed. Am. Chem. Soc. Exp. Biol.*, **17**, 1075 (1958).
- T.C. Laurent and J.R.E. Fraser, *FASEB J.*, **6**, 2397 (1992).
- B.P. Toole, in E.D. Hay ed. *Cell Biology of Extracellular Matrix*, Plenum, New York, 259 (1981).
- M. Yoneda, S. Shimizu, Y. Nishi, M. Yamagata, S. Suzuki, and K. Kimata, *J. Cell. Sci.*, **90**, 275 (1988).
- I. Ellis, A.M. Grey, A.M. Schor, and S.L. Schor, *J. Cell. Sci.*, **102**, 447 (1992).
- R.N. Feinberg and D.C. Beebe, *Science*, **220**, 1177 (1983).
- D.C. West, I.N. Hampson, F. Arnold, and S. Kumar, *Science*, **228**, 1324 (1985).
- D.C. West and S. Kumar, in *The Biology of Hyaluronan*, Ciba Foundation symposium 143, Wiley, Chichester, England, 187 (1989).
- S.D. Banerjee and B.P. Toole, *J. Cell. Biol.*, **119**, 643 (1992).
- Y. Nakahara and T. Ogawa, *Tetrahedron Lett.*, **28**, 2731 (1987); *Carbohydr. Res.*, **173**, 306 (1988).
- G. Grundler and R.R. Schmidt, *Carbohydr. Res.*, **135**, 203 (1985).
- Physical data for new compounds are given below, values for [α]_D and δ_H were measured at 25°C for solutions in CH₂Cl₂ and CDCl₃ respectively, unless noted otherwise. Signal assignment such as 1³ stands for a proton at C-1 of monosaccharide residue 3. **1**: [α]_D -45°

(c 1.0, H₂O); R_F 0.55 in 4:2:2:1 nBuOH-EtOH-H₂O-AcOH; δ_H (D₂O) 2.026 and 2.046 (2s, 2Ac), 3.810 (s, OMe), 4.491, 4.541, and 4.591 (3d, 7.8-8.4 Hz, 1², 1³, and 1⁴), 5.015 (d, 8.6 Hz, 1¹); FAB MS m/z 883 (M+H). 3: [α]_D +80° (c 1.0) (α:β 3:2); δ_H 1.789, 1.912, and 1.978 (3s, 3Ac), 2.209 (s, Lev), 2.327 and 2.379 (2s, 2MeBz), 5.585 (d, 8.1 Hz, 1²), 6.600 (d, 3.7 Hz, 1¹α), 6.631 (d, 8.8 Hz, 1¹β). 4: [α]_D +31° (c 1.0); δ_H 1.239 and 1.250 (2s, CMe₂), 2.160 (s, Lev), 2.316 and 2.370 (2s, 2MeBz), 3.678 (s, OMe), 5.048 (d, 7.5 Hz, 1¹), 5.287 (d, 8.2 Hz, 1²). 8: [α]_D +57° (c 1.0); δ_H 1.793, 1.918, and 1.965 (3s, 3Ac), 2.185 (s, Lev), 2.355 and 2.389 (2s, 2MeBz), 3.706 (s, OMe), 5.027 (d, 7.7 Hz, 1¹), 5.514 (d, 8.4 Hz, 1²). 9: [α]_D +25° (c 1.0); δ_H 1.224 and 1.254 (2s, CMe₂), 2.188 (s, Lev), 2.338 and 2.390 (2s, 2MeBz), 3.695 (s, OMe), 5.026 (d, 7.5 Hz, 1¹), 5.408 (d, 8.0 Hz, 1²). 10: [α]_D +70° (c 1.0); δ_H 1.768, 1.889, and 1.923 (3s, 3Ac), 2.162 and 2.166 (2s, 2Lev), 2.305, 2.341 x 2, and 2.380 (3s, 4MeBz), 3.683 (s, OMe), 4.443 and 4.957 (2d, 7.2 and 7.9 Hz, respectively, 1¹ and 1³), 5.025 and 5.414 (2d, 8.3 Hz, 1² and 1⁴). 11: [α]_D +59° (c 1.0); δ_H 1.779, 1.836, 1.882, 1.913, and 1.946 (5s, 5Ac), 2.337 x 2, and 2.362 x 2 (2s, 4MeBz), 3.690 (s, OMe), 4.504 and 4.990 (2d, 7.2 and 7.7 Hz, respectively, 1¹ and 1³), 5.149 and 5.458 (2d, 8.4 Hz, 1² and 1⁴). 12: [α]_D +6° (c 1.0); R_F 0.48 in 10:9:1 CH₂Cl₂-acetone-HOAc; δ_H 1.757, 1.824, 1.842, and 1.881 x 2 (4s, 5Ac), 2.298, 2.333, 2.344, and 2.382 (4s, 4MeBz), 3.689 (s, OMe), 3.783 and 3.859 (2d, 9.5 and 9.0 Hz, respectively, 5¹ and 5³), 4.438 and 5.057 (2d, 7.5 and 7.7 Hz, respectively, 1¹ and 1³), 5.370 and 5.567 (2d, 8.3 and 8.4 Hz, respectively, 1² and 1⁴). The corresponding dimethylester was prepared by treatment with CH₂N₂: δ_H 1.767 x 2, 1.884, 1.894, and 1.908 (4s, 5Ac), 2.308, 2.340 x 2, and 2.385 (3s, 4MeBz), 3.386, 3.577, and 3.686 (3s, 3OMe), 3.722 and 3.794 (2d, 9.7 and 8.9 Hz, respectively, 5¹ and 5³), 4.355 and 5.002 (2d, 7.6 and 7.0 Hz, respectively, 1¹ and 1³), 4.971 and 5.285 (2d, 8.4 and 8.3 Hz, respectively, 1² and 1⁴).

- 14 T. Fukuyama, A.A. Laird, and L.M. Hotchkiss, *Tetrahedron Lett.*, **26**, 6291 (1985).
- 15 R.R. Schmidt, J. Michel, and M. Roos, *Liebigs Ann Chem.*, 1343 (1984).
- 16 H. Kunz and H. Waldman, *Angew. Chem.*, **96**, 49 (1984).
- 17 Y. Hayakawa, H. Kato, M. Uchiyama, H. Kajino, and R. Noyori, *J. Org. Chem.*, **51**, 2400 (1986).
- 18 J.H. van Boom and P.M.J. Burgers, *Tetrahedron Lett.*, 4875 (1976).
- 19 N. Jeker and C. Tamm, *Helv. Chim. Acta*, **71**, 1895, 1904 (1988).
- 20 K. Omura and D. Swern, *Tetrahedron*, **43**, 1651 (1978).
- 21 B.O. Lindgren and T. Nilsson, *Acta Chem. Scand.*, **27**, 888 (1973).
- 22 B. Kraus and B. Roth, *J. Org. Chem.*, **45**, 4825 (1980).
- 23 M.S. Motawai, J. Wengel, A.E.S. Abdel-Megid, and E.B. Pedersen, *Synthesis*, 384 (1989).

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